Case Report



is detected, before a definitive diagnosis, the hematological

parameters must be re-evaluated after iron supplementation. Here

is a case of nine months old child with recurrent hemolytic anemia

A seven months old female child was referred to our hospital

with fever. She looked very pale since one month. On examination,

there was amid hepatomegaly. Laboratory test results can be seen

on (Tables 1 & 2). On peripheral blood film examination, RBCs

were dominantly microcytic hypochromic revealing high degree

of anisopoikilocytosis [2]. There were teardrop cells, normoblasts,

and polychromatic red cells (Figure 1). A clinical diagnosis of

hemoglobinopathy was made. Before blood transfusion, an

EDTA blood samples was sent to the laboratory for hemoglobin

electrophoresis (Figure 2). All the hemoglobin fractions are within

normal limit compared to age-specific manufacturer's reference

range. Because iron deficiency was already excluded, patient was

suggested to repeat the hemoglobin electrophoresis when she is

already one year old. Blood transfusion was given gradually.

and clinically suspected of having thalassemia.

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Challenges in Early Infant Diagnosis of Beta Thalassemias



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Case Report

Introduction

Thalassemias are a heterogeneous group of genetic disorders that result from a reduced rate of synthesis of or ß chains. The genetic defects of hemoglobin are the most genetic disorders worldwide. They occur in tropical and subtropical areas and most appear to have been selected because the carrier state affords some protection against malaria. Indonesia is among a group of countries with high prevalence of thalassemias. The burden of this disorder in the country is of such a magnitude that it represents a major public health concern. Laboratory diagnosis of thalassemia, which consists of blood film examinations and hemoglobin electrophoresis, are the most common modalities in our country since DNA analysis for or ß globin mutations is high cost and only available in the capital city. On the other hand, diagnosis of thalassemia becomes more complicated because of high prevalence of iron deficiency anemia in the country [1]. Health survey data in 2001 showed the prevalence of deficiency anemia in infants 0-6 months, infants 6-12 months, and children under five years old were 61.3%, 64.8%, and 48.1%, respectively. Iron deficiency should be excluded because it causes falsely low HbA2 fractions on hemoglobin electrophoresis using micro capillary electrophoresis method. Whenever iron deficiency

<u>le 1</u> : Complete blood count result of the patient.					
Parameters	Results	Unit	Reference Range		
Hemoglobin	3.7	g/dL	10.1-12.9		
Erythrocyte count	0,9	10^6/ L	3.2-5.2		
Hematocrit	6.5	%	32-44		
MCV	70.7	fL	73-109		
MCH	40.2	pg	21-33		
МСНС	56.9	g/dL	26-34		
RDW-CV	30.6	%	11.5-14.5		
Leukocyte count	13980	/ L	6000-17500		

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Thrombocyte count	376000	/ L	150000-450000
Serum iron	53.15	g/dL	33-193
TIBC	194.00	g/dL	228-428
UIBC	140.60	g/dL	110-370

<u>Table 2</u>: Complete blood count results of both parents.

Parameters	Father, 41 years old	Mother, 33 years old	Unit
Hemoglobin	11.8	13.5	g/dL
Erythrocyte count	6.26	6.91	10^6/ L
Hematocrit	38.6	41.2	%
MCV	61.7	59.6	fL
МСН	18.8	19.5	pg
МСНС	30.6	32.8	g/dL
RDW-CV	17.4	17.8	%
Leukocyte count	7610	10020	/ L
Thrombocyte count	286000	310000	/ L

Note: The hemoglobin electrophoresis results of both parents can be seen below.

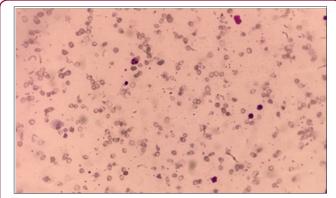


Figure 1: Peripheral blood film.

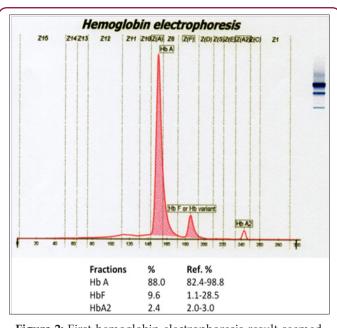


Figure 2: First hemoglobin electrophoresis result seemed normal.

Before discharge, hemoglobin level was 11.3 g/dL. Unexpectedly, a month later, she came again to the emergency department with fever. She looked very weak and pale. Laboratory examinations showed hemoglobin level 7.5 g/dL. Before blood transfusion, an EDTA blood sample was sent to the laboratory for hemoglobin electrophoresis (Figure 3). The electrophoresis was, again, within normal limit according to manufacturer's reference range [3]. Because of high suspicion of thalassemia and after excluding other causes of hemolytic anemias, we suggested both parents to have hemoglobinopathy screenings (Figure 4). Hematology results of them can be seen below.

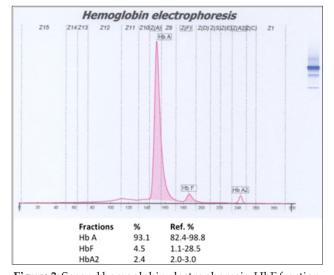
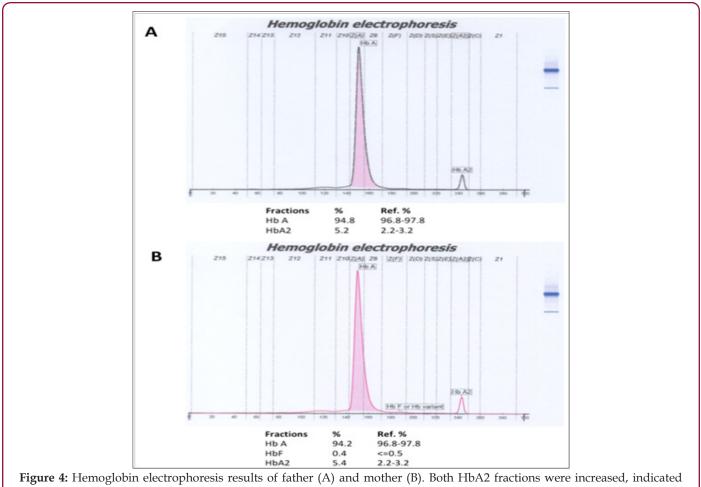


Figure 3: Second hemoglobin electrophoresis. HbF fraction was decreased compared to the first result.

Discussion

Thalassemias can be defined as ß-thalassemias depending on the defective globins chain and the underlying molecular defects. In patients with ß-thalassemia major, clinical presentations usually occurs between 6 and 24 months of life, with severe microcytic, mild jaundice, and hepatosplenomegaly. Hematological diagnosis is based on reduced hemoglobin level (<7 g/dL) and very low MCV. The peripheral blood smear shows severe erythrocyte morphologic changes with marked poikilocytosis (speculated tear-drop cells), target cells, and numerous nucleated red blood cells (NRBC). The number of NRBC is related to the degree of ineffective erythropoietin.

Additional tests are required to exclude iron deficiency anemia. After having complete hematological test, quantitative HbA2 determination is the most valuable test for thalassemia detection. Patients should receive iron supplementation before having hemoglobin electrophoresis test because iron deficiency can falsely lower the HbA2 level.



them as ß-thalassemia carriers.

The expected normal range for HbA2 is between 2.4 and 3.2% in normal subject, while in typical ß-thalassemia carriers, it is between 3.6 and 7%. Values between 3.2 and 3.6% are considered borderline and they need further investigation, especially in young subjects or couple at risk. The normal range for HbF in adult life is usually <1.5% of total hemoglobin. In the prenatal period, the synthesis of hemoglobin switches from fetal (HbF) to that of adult (HbA). Factors that control the switching at the molecular or cellular level are as yet unknown. High levels of HbF in adults are known to be associated with several genetic disorders of goblin chain synthesis, particularly hereditary persistence of fetal hemoglobin (HPFH), thalassemia, and several combinations of thalassemias to other abnormal hemoglobin's. Information on the developmental changes of HbF levels in tha above conditions during the neonatal period and early infancy is limited. This causes difficulties in establishing the diagnosis of ß-thalassemia heterozygotes at this age, as happened in this patient.

Interpreting hemoglobin electrophoresis result of early infants are challenging because there are no established reference range for HbF for this age group. In this patient, compared to previous result, the decrease of HbF fraction on second test was most likely caused by blood transfusion one month ago, and was not a natural occurrence. Because the molecular test is not feasible in our region, we suggested the parents to have thalassemia screening. Both of them were classified as ß-thalassemia carriers because they have low hemoglobin levels, low MCVs, low MCHs, and increased HbF fraction on electrophoresis. Beta-thalassemias are recessive trait. Thus, the clinical relevant phenotypes result from homozygosis or double heterozygosis for different globing gene defects. They can also manifest from co-inheritance of thalassemia trait. Definitive diagnosis of thalassemia for this patient will be done at older age. Blood transfusion should not be done prior to the test to prevent misdiagnosis.

Conclusion

Thalassaemia is the leading genetic problem in the Southeast Asia population. A proper laboratory diagnosis is crucial for characterizing the different forms of thalassemia with important implications for diagnosis, prevention, and treatment. The iron status should always be taken into account when evaluating individuals screened for thalassemia. Diagnosis of ß-thalassemia using capillary electrophoresis in neonates and early infants are challenging because there is no exact reference ranges for this age group. In such cases when there is high suspicion of thalassemia but inconclusive hemoglobin electrophoresis result, it is suggested that the parents should have hemoglobinopathy screening.

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