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The Effect of Granulocyte Colony Stimulating Factors (GCSF) on Expansion of CD34⁺ Hematopoietic Stem Cells



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Abstract

Stem cells are alternatives therapeutic treatment, mainly for diseases that cannot be treated by medical or surgical treatment. CD34+ hematopoietic stem cells which is isolated from autologous peripheral blood cell is a good source because it's lowering immune rejection However, the limit volume of peripheral blood can be taken from a patient, limit the total number of CD34+ cell and therefore will be inadequate for therapeutic efficacy. In the previous addition of GCSF, TPO and SCF to CD34+ culture increased the number of CD34+ cell. But the addition of several cytokines will increase the cost of culture become a burden for patients who are seeking for stem cell therapy. In this study we examined the effect of GCSF on the expansion of CD34+ cell. For that purpose, we isolated and expanded CD34+ cell from peripheral blood using GCSF. As a control cultures, we used GCSF, TPO and SCF addition. Both culture was incubated for 10 days, and then counted the number of CD34+ cells. The results showed the amount of CD34+ cells can be increased only with GCSF. When compared to the control group, the number of CD34+ cells treated with GCSF were lower than which are an average 51,250 cells compare to 3,300,000 cells, respectively. There is a significant difference between the number of cd34 cells treated only with GCSF and the control group (p=0.04). Furthermore, the differentiation potential of CD34+ /GCSF expanded cells should be performed. In conclusion, GCSF alone has positive effect in increasing CD34+ cells number although the increase is lower compared to if all the treatment is combined.

Keywords: CD34+ cell; GCSF; Hematopoietic cell; Peripheral blood

Abbreviations: GCSF: Granulocyte Colony Stimulating Factors; CSF: Colony Stimulating Factors; HSC: Hematopoietic Stem Cells

Introduction

Human tissue has varying levels in regeneration capacity, even some of musculoskeletal tissues such as the cartilage and ligament have no ability to regenerate at all. In some condition when that tissues had trauma or diseased, they cannot regenerate. Until now, medical therapy has not been able to make growth or produce new tissue that tissue disorders are often end with surgical therapy such as replacement surgery. The solution is to make new tissue by regeneration therapy. Tissue engineering technique using stem cell could be the promising alternative therapy because of its ability to regenerate new tissue. Stem cells are cells that have ability to renew and multiply themselves and form cells and tissues structuring the organism body. Stem cells have some superior characteristics, namely plasticity produce regenerative tissue and prevent degeneration.

Stem cells can be categorized into embryonic stem cells and tissue stem cells. There are some limitations in using them such as ethical problem and teratoma risk. Tissues stem cells found in some tissues or organs. There are 2 types of tissue stem cells, mesenchymal stem cells and hematopoietic stem cells. Mesenchyme stem cells are potential sources of tissue engineering. However, the application of mesenchyme stem cells faces some limitations in sampling process and its nature. The sampling process causes pain and complication such as infection and sepsis. The other limitations are the characteristic of mesenchyme stem cells such as the number of cells, range of age, proliferation and its differentiation related to age of the donor. Hematopoietic stem cells are favorable because its characteristic, namely easy taking, low complication risk, and have differentiated potential without being affected by the age of the donor.

Hematopoietic stem cells are progenitor cells forming blood cells-lymphoid cells and myeloid cells. Cells, that come from bone marrow and blood respectively. Hematopoietic stem cells have pluripotent and plasticity so that they can form nonhematopoietic cells. The change of lineage is assumed to undergo some mechanism namely trans differentiation, dedifferentiation, and cell fusion. Hematopoietic stem cells can move toward the target tissues and form new cells and tissue. Major source of HSC is CD34* cells, the other is CD133. Today, Hematopoietic Stem Cells (HSC) are clinically efficiently used for blood cancer therapy. More research indicates other uses of HSC for non-hematopoietic tissue such as for musculoskeletal tissue especially for ischemic chronic therapy in legs and osteogenesis imperfecta. Some researchers reported the use of hematopoietic stem cells for regeneration of non-hematopoietic tissue in experimental animals. Matsumoto reported the healing of rat's femur bone fracture, through vasculogenic and osteogenic capabilities of CD34+ stem cells. Shi et al. reported the capability of CD133+ stem cells for the muscle tissue regeneration by healing of muscle injury in rats. Terayama reported the healing of osteonecrosis of rats' hip joint. Basuki reported the healing of cartilage defect of the rat's knee joint using CD34⁺ stem cell. It showed that CD34⁺ stem cell has capability of cartilage regeneration. These result indicate the differentiation potential that CD34+ cells have differentiation potential to become endothelial and osteoblasts cells.

CD34* stem cell can be obtained by doing selection process of MNCs cells population. MNCs cells population taken by isolating the human body tissue such as solid or liquid. Skin and fat are examples from solid human body tissue whereas bone marrow, peripheral blood and cord blood are examples from liquid human body tissue. The process of isolating MNC cells from body liquids more preferable because of the easier process than isolating from solid human tissue. The collection process from bone marrow fluid and umbilical cord blood is inconveniences thus increased morbidity and easily contaminated. Therefore, collection process from peripheral blood human is safer and more comfortable for patients with some limitation [1]. The limitation of obtaining CD34* cells from peripheral blood is the number of cells that can be used is not enough for therapeutically purpose [2-4].

The challenge is how to increase the number of CD34* cells from peripheral blood selection for stem cell therapy. We need effective method to increase it. The previously methods are mobilization CD34* from the bone marrow using cell growth factors namely GCSF (Granulocyte Colony Stimulating Factors) [5,6]. Colony Stimulating Factors (CSF), is hematopoietic growth factor, which plays a role in the regulation of bone marrow cell mobilization, proliferation and differentiation [7]. In fact, this method is done by give an injection to patient that feels uncomfortable to patient, another method is expansion CD34* cells (*In vitro*) in a medium containing cell growth factors. The standard method of CD34* cells expansion is cell expansion with multiple or combination of several cell growth factors, namely GCSF, SCF, TPO. GCSF has important role in CD34* cells expansion [2,4,7,8]. CD34* cells expansion with a mixture of several growth factors is less economical. the alternative

is to use only one growth factor, namely GCSF. The purpose of the study was to determine the effectiveness of administration GCSF only compared with the administration of GCSF mixed with SCF and TPO.

Material & Methods

Ethics Statement

Research was done at Stem Cell Research and Tissue Engineering Centre, Medical Faculty of University Pembangunan Nasional Veteran, Jakarta, Indonesia. All procedures were approved by Ethical Committee of University Pembangunan Nasional Veteran, Jakarta, Indonesia. Blood donor was volunteer who has been explained all of the procedures and use of their blood samples in this study and has knowingly signed the informed consent.

Blood Collection

Human peripheral blood was collected from 2 males, 30 years, healthy donors. Donors had no history of hepatitis, HIV, malignancy, or bone marrow disease and never had chemo or radiotherapy. Donors signed informed consent and undergone a preliminary blood examination to determine infection risk of Hepatitis and HIV, and both results were negative. All procedures in taking and examination of blood samples were conducted in Al Fauzan Hospital, Jakarta, Indonesia. About 100 ml peripheral blood was collected.

Peripheral Blood CD34⁺ Cells Selection

All collected blood were isolated with Ficoll gradient centrifugation and selected to CD34+ using Mini Macs column system. Blood specimens were diluted with PBS+ KCl solution, filtrated with Ficoll and centrifuged. Buffy coat layers were taken, washed and supernatant was removed until only MNC cells were available to be counted and viability checked. Afterward we used MACS isolation CD34* kit (Militenyi) for CD34* selection. MNC cells were washed and labeled by adding 300 µl buffer solution (auto MACS TM Rinsing Solution and MACS BSA Stock Solution, 20:1), FcR Blocking 100 μ l, and CD34* micro-beads 100 μ l, and the suspension was incubated for 30 minutes at 2-8°C. Subsequently, 10 mL buffer solution was added to cell suspensions and centrifuged. The supernatant was removed, and cells were re-suspended with 500 L buffer. Cells were separated with separator column. About 500 L of buffer was added into the column along with the cell suspension. The column was washed with 500 L buffer solution three times. CD34⁺ cells retained in the column were pushed with a syringe into a tube. Cell suspensions were centrifuged, the supernatant was removed, and cells were counted amount of cell and its viability.

CD34⁺ Cells Expansion

We prepared 24 well plates, 12 well plates for experimental group and 12 well plates for control group. We prepared two type of cell culture medium. Medium $1:10\,4\,\text{CD34}^+$ cells, GCSF (100 ng/ml, Miltenyi), Stem line II Hematopoietic Stem Cell Expansion medium (Sigma). Medium $2:10\,4\,\text{CD34}^+$ cells, in GCSF (100ng/ml, Miltenyi), TPO (100ng/ml, Miltenyi), SCF medium (100ng/ml, Miltenyi), Stem line II Hematopoietic Stem Cell Expansion medium (Sigma). We

put medium in well plate: medium 1 for experimental group and medium 2 for control group 1 (exin 12 well plate. We observed it for 10 days, and $CD34^{\circ}$ cell was counted in day 10^{th} .

About 10.000 CD34* cells were planted in each well from 24 well plates using the stem line medium containing GCSF cytokines. As a control the Stem line medium was added to the GCSF, SCF and TPO cytokines. The dose of GCSF used was 100 ng/ml for each group and repeated 4 times [3,9]. For control it is used a medium that added with GCSF(100 ng/ml), SCF (100 ng/ml) and TPO (100ng/ml). Same as with the addition of GCSF, then the control had 4 times repetition too. The cultures were observed for 10 days and then counted for the amount of cells.

Statistical Analysis

Data were descriptively and comparatively analyzed by SPSS version 17.0 and expressed as mean and SD. The comparative analysis from amount of CD34 $^{+}$ cells were evaluated using t test. P, 0.05 was considered statistically significant [10].

Result & Discussion

Peripheral Blood CD 34 Cell Selection

The result of donor blood examination was 13.9 Hb level of g/dl, leucocyte count of 7.4×10^3 / mm, and platelet count of 209×10^3 / L. The amount of CD34 $^+$ cells : 3.7×10^6 cells, with 95 % viability.

CD34⁺ Cells Expansion

In the experimental groups (GCSF medium) and the control groups (GCSF,SCF,TPO medium), the expansion of CD34 $^{\circ}$ cells were occurred well without any contamination. According to observation until the 10 th days, the cell form was maintained (like previous form) and didn't attach to the plastic. The number of cells continued to increase but on the 10 th day the increase of CD34 $^{\circ}$ cells is more common in the control group. On the 10 th day, in the experimental groups amount of CD34 $^{\circ}$ cells are 5.125 x 10 $^{\circ}$ while at the control groups amount of CD34 $^{\circ}$ cells are 330 x 104 (Table 1) and (Figure 1). There was significant difference (t test, p = 0.04) in amount of CD34 $^{\circ}$ cell produced from cultures using GCSF alone with cultures using a combination of GCSF, TPO, and SCF.

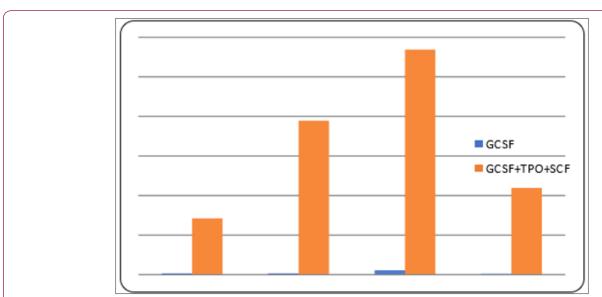


Figure 1: Comparison between amount of CD34⁺ cells in experimental group and control group on the 1st day and 10th day, in every repetition.

Table 1: Comparison between amount of CD34⁺ cells in experimental group and control group on the 1st day and 10th day.

Group	Day 1	Day 10
Experimental Group (GCSF medium)	1×10^4 cells	5.125 x 10 ⁴ cells
Control Group (GCSF, SCF, TPO medium)	1 x 10 ⁴ cells	330,0 x 10 ⁴ cells

This result showed that the cell proliferation with GCSF alone give lower cells proliferation compare to cells treated with combination of GCSF, TPO, and SCF. This result was in accordance with the statement that TPO and SCF cytokines can increase the number of peripheral blood progenitor cells and work in synergy with the GCSF [2,11,12].

Figure 2 showed that the amount of CD34 $^{\circ}$ cells continued to increase both in control and experimental group from the 1 st day until the 10 th day. However, this result hasn't shown how

the differentiation of CD34 * cell yet. For this reason, the study is still continuing with characterization phase. It is hoped that the characterization results will show whether the CD34 * cells are still hematopoietic cells, or it can differentiate to mesenchymal cells too. There are some limitations in this study such as time and amount of the cells. The time for the expansion of stem cells in this study is only 10 days, which is not enough to expand more stem cells. We suggest for the next study, the time for expanding the stem cell should be prolonged and the amount of cells should be increased. We need further research for solve this questions.

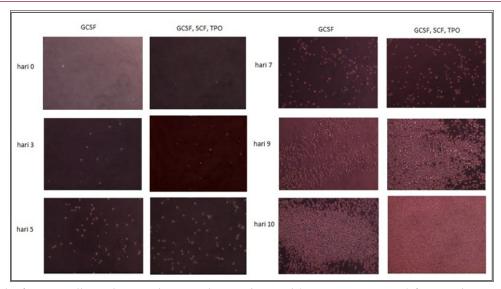


Figure 2: Growth of CD34+ cell in culture with GCSF alone and control (GCSF + TPO + SCF) from 1st day until 10th day.

Conclusion

GCSF has positive effect on CD34⁺ cells expansion in vitro but the effect is less than the combination of GCSF, SCF and TPO.

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