

# Isolation of Salmonella from Goats Slaughtered at Abyssinia Export Abattoirs

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## ARTICLE INFO

**Received:** 📅 June 09, 2023

**Published:** 📅 June 19, 2023

**Citation:** Ibrahim Sadik, Megersa Bedassa and Ephrem Shimelis. Isolation of Salmonella from Goats Slaughtered at Abyssinia Export Abattoirs. Biomed J Sci & Tech Res 51(1)-2023. BJSTR. MS.ID.008053.

## ABSTRACT

A cross-sectional study was conducted between February 2020 and March 2020 on apparently healthy slaughtered goats at Abyssinia export abattoirs located at Bishoftu towns to estimate the prevalence of salmonellosis from goat meat by the isolation and identification conventional tests. A total of 200 samples carcass swab (60), hand swab (40), knife swab (40) and faeces (60) were collected from goats slaughtered at Abyssinia export abattoir. The samples were examined for the presence of Salmonella by the conventional techniques of Selective media and biochemical tests for isolation salmonella species. From the total of 200 animals examined, 5 (2.5%) were positive of which 4 (2%) were young and 1 (0.5%) were adult. Statistically significant difference ( $p=0.01$ ) in the prevalence of Salmonella was observed between the ages of goats. Of the total 200 samples examined from apparently healthy slaughtered goats, 5 (2.5%) samples were Salmonella positive. Salmonella was isolated from 1.5% carcass swab, 0% hand swab, 0.5% knife swab and 0.5% faeces samples. However, there was a significant difference between samples ( $P=0.01$ ). The results of this study showed the potential risk of goats as sources of pathogen for humans in the study area. These findings stressed the need for implementation of preventing close contact of offal and carcasses during evisceration.

**Keywords:** Abattoir; Abyssinia; Carcass; Goats; Isolation; Salmonella

**Abbreviations:** AAU: Addis Ababa University; CVMA: Collage of Veterinary medicine and agriculture; IBC: Institute of Biodiversity Conservation; ISO: International Organization for Standardization; MASL: Meter Above Sea Level; MR: Methyl Red; OIE: Office International Des Epizootics; PWB: Peptone water broth; RVS: Rappaport-Vassiliadis Soy Peptone; TSI: Triple Sugar Iron; VP: Voges Proskauer; XLD: Xylose Lysine Deoxycholate

## Introduction

Ethiopia consist a diversity of livestock population and possess the large livestock population from the African countries (IBC, 2007). The goat population is estimated to be around 30.20 million heads.in which 1,128,000 goats were estimated to be consumed domestically (CSA [1]). Livestock productions plays an important role to smallholder farmers and the national economy of the country in generating income to farmers, creating job opportunities, ensuring food security, providing services, contributing to asset, social, cultural and environmental values, and sustain livelihoods (Behnke, et al. [2,3]). However, the sector is affected by infectious diseases (viral, bacterial, parasitic and fungal diseases), feed shortage and lack of veterinary service

(Ganteby [4]). From bacterial disease, food borne zoonotic pathogenic disease such as Salmonella an importantly affect both human and animals in developing countries including Ethiopia. Salmonellosis is one of the widest spread foods borne zoonoses with the main cause of mortality and morbidity to humans and animals (D'Aoust [5]). Salmonellosis is a disease, or a group of diseases caused by a wide variety of Salmonella serovars in various hosts including goat (OIE Manual [6]) which remains a serious problem with public health significance throughout the world (Tabaraie, et al. [7]).

Salmonella are Gram negative, bacillus shaped, non-spore forming, aerobic and facultatively anaerobic organisms belong under family Enterobacteriaceae, were most of them have peritrichous flagella

that use for motility (Freeman, et al. [6,8,9]). They have two species such as, *Salmonella enterica* and *Salmonella bongori*. *Salmonella enterica* has six subdivisions of *S. enterica* subspecies 1-6 exist with over 2500 serovars currently identified and several common serovars to human clinical infections (Dworkin, et al. [10]). The main forms of salmonellosis happen are in three forms which is peracute septicaemia, acute enteritis, and chronic enteritis. Common clinical signs of salmonellosis are abortion, upset stomach, diarrhea, fever, and pain and also cramping in your belly. Although, the clinical signs may vary from species to species (Blood, et al. [11]). They are facultative intracellular pathogens that cause disease in animals, they are commonly asymptomatic carriers. *Salmonella* was a highly concentrated intensive study on cattle and sheep; however, there is little published information on the conveying of *Salmonella* in goats, although goat meat was one of a source of *Salmonella* food poisoning in Ethiopia (Kadaka, et al. [12]).

The diagnosed salmonellosis can be based on taking history and laboratory examination of the *Salmonellosis* using feces, carcass swab, feed and water (Carlton [13]). However, identification *salmonella* is the most difficult at clinical sign because the bacteria may be intermittently, and it may be sample taking is important for examine. However, for identification of the bacteria at least requires five days for use of standard cultural techniques (House, et al. [14]). Thus, cultural technical for isolation of salmonellosis was Pre-enrichment, enrichment, plating out and different biochemical and serological test according to the set of standards (Pieskus [15]). The Ethiopia goat population was suffering from a contagious disease like *Salmonellosis* and *Brucellosis*. *Salmonella* is the main cause of meat contamination that can affect animal and human being (Pepin, et al. [16,17]). The risk factors for the exposure of salmonellosis contamination of animal carcasses and fresh meat during slaughter and dressing by the faeces, the skin, water, air, intestinal contents, processing equipment and humans, and can be transferred to the carcass during skin removal and evisceration (Hansson, et al. [18,19]). Therefore, this study was designed to isolation of *salmonella* from different type of samples (knife swab, hand swab, carcass swab and fecal) collected from goats slaughtered in Abyssinia export abattoir.

## Materials and Method

### Study Area

The study was conducted at Abyssinia export abattoirs in Bishoftu town, East Showa zone of Oromia regional state. Bishoftu town was located at 9°N latitude and 40°E longitude at altitude of 1850 masl with the annual average temperature ranges of 12.3 °C to 27.7 °C with an overall average of 18.7°C. In thus, area the climate and soil type are the same with those in the highland areas in Ethiopia. It got heavy rain in the season of June to September with an annual rain fall of 866 mm and which is 84% got in that long rainy season. However, there are many livestock farms of beef and dairy cattle, small ruminant, poultry and

equines and there are many modern slaughter services (abattoirs) in the city. From these abattoirs, Ethiopia is exporting millions of tons of meat of cattle, sheep and goats to Middle East countries and gain good external currency. Hence, this was designed to undertake salmonella isolation from goats brought to Abyssinia Slaughtering Service House for slaughter.

### Study Design and Animals

The cross-sectional study was conducted on isolation and identification of *salmonella* from apparently healthy goats slaughtered at Abyssinia Export abattoir service from February 2020 to March 2020. Conveniently samples were collected from 200 goats purchased from different zones of Oromia region (Arsi, Hararghe, Bale and Borena). The animal was grouped into two classes as young, and adult based on dentition by Vatta [20] where the animal which have only one pair permanent of incisors teeth were considered as young (<1 year to 2 years) while the goats which have two or more pairs of permanent incisors teeth were considered as adult (>2 years).

### Sample Collection and Transportation

Based on its accessibility of samples during the sample collection, the samples were collected from 200 goats provided for slaughtering at Abyssinia export abattoir. The samples were collected aseptically from goats during slaughtering operation. Thus, sample such as: feces, carcass swab, knife swab and hand swab were collected aseptically using sterile cotton swab for organ samples and put into sterile universal bottles with peptone water media for transportation according to the recommendation (Quinn, et al. [21]). Each sample was collected from each slaughtered goat during the operation. Because of fasting, abattoir worker does not take four samples at the same time from the same goat. All samples were labeled apparent identifying species of the animal, age and type of sample and sampling ID number. After putting the samples in an ice box with ice pack. The sample was transported immediately to AAU, CVMA veterinary public health laboratory for bacteriological processing.

### Isolation *Salmonella*

**Sample Preparation and Inoculation on Nonselective Pre-Enrichment:** The fresh feces sample of 10 g was broken into pieces by using Mortar and pestle and homogenized by homogenizer before transferred to pre-enriched in 90 ml of buffered peptone water (BPW). also. the swab samples were transferred to pre-enrichment of buffered peptone water in the ratio of 1:9 BPW (HI Media M1494, Mubi, India). Both the Swab and fecal sample were incubated for 24 hr at 37°C.

**Culturing *Salmonella* on Selective Media:** On the second day, 0.1 ml (100 uL) of the pre-enrichment broth will be transferred to 10 ml Rappaport-Vassiliadis soy peptone (RVS) and incubated at 37°C overnight (18-24 hours). On the third day, the suspension culture of





**Annex Figure 1:** Salmonella colony morphology and its biochemical characteristics.

Note: The name of picture: XLD colony, TSI test, Simmon citrate test, MR test, VP test and Indol test, respectively



## Result

This study was conducted to isolate salmonella species from goats slaughtered at Abyssinia export abattoir during February 2020 to March 2020 to isolate salmonella species. A total of 200 samples from Abyssinia export abattoirs (60 carcass swab, 40 hand swab, 40 knife swab and 60 feces) were collected and examined for the presence of salmonella species. Among these five salmonella species was isolated of showing characteristics salmonella, i.e., the colony appears as translucent with the black center and hydrogen sulfide production from thiosulfate is easily detected because colonies become dark or gloomy, due to the precipitation of ferric sulfide on XLD agar and positive on biochemical tests for salmonella and. The Salmonella was isolated from all sample types except hand swab. The prevalence varied with sample site and time of sampling. When compare the prevalence of salmonella by the sample sources, the salmonella was more prevalent on carcasses (1.5%). In the case of age, it's more prevalent in young (2%) than adults.

### Prevalence of Salmonella from Different Sample Sources

Salmonella was isolated from the samples collected from Carcass swab 3(1.5%), Hand swab 0(0%), Knife swab 1(0.5%) and Feces 1(0.5%) as it isolated and identified on the Selective media and Biochemical analysis of the sample. There was significant difference ( $P < 0.05$ ) in the frequency of Salmonella isolation among the samples (Table 1).

**Table 1:** The prevalence of salmonella from different sample sources.

Types of samples	Examined	Positive	Percentage in each sample	Percentage in total	$\chi^2$	p-value
Carcass swab	60	3	5%	1.50%	6.4431	0.011
Hand swab	40	0	0%	0%		
Knife swab	40	1	2.50%	0.50%		
Feces	60	1	1.67%	0.50%		
Total	200	5	2.50%	2.50%		

### Prevalence of Salmonella Between the Age of Goats

The prevalence of salmonella was high in young goats when compared with that of adult goats (2% and 0.5%, respectively) (Table 2). The prevalence of salmonella was different between the age of the goats from which the sample was collected. There was significant difference ( $P < 0.05$ ) in the frequency of Salmonella isolation between adult and young (Table 2).

**Table 2:** Prevalence of salmonella between the age of goats slaughtered at selected abattoir.

Age	Examined	Positive	Prevalence of age%	Prevalence from total%	$\chi^2$	p-value
Adult	70	1	1.40%	0.50%	6.4431	0.011
Young	130	4	3.10%	2%		
Total	200	5	2.50%	2.50%		

## Discussion

In the present study the carrier state of salmonella in apparently healthy goats slaughtered at Abyssinia export abattoir was 2.5%. Among the total of 200 animals examined, 5 (2.5%) were positive for Salmonella of which 1(0.5%) were adult and 4 (2%) were young. The prevalence of Salmonella was higher in young people than adults. This difference was statistically significant ( $P=0.01$ ). This variation in prevalence of Salmonella might be due to management differences and handling of owner in this age (Wassie [22]). The higher prevalence in young might be due to higher Salmonella carrier rate in the study population. The prevalence of Salmonella in apparently healthy slaughtered goats in this study was 2.5%. This result is consistent with the range cited by (D'Aoust [23]) which indicated that the prevalence of Salmonella in goats falls between 1 to 18.8%. Also, the result obtained was agreed with the result of Ferede, et al. [24] done on the distribution of salmonella was higher in young than adults that was 2%. However, this result is the opposite of the previous reports. This variation in result may be due to the young congregating with your mother and cross contamination during sampling or due to different bacteriological procedures followed. In the present study, Salmonella was isolated in 1.28% of the abdominal muscle of goat. Isolation of salmonella from goat carcass swab was 17.7% in Dire Dawa municipal abattoir in Ethiopia.

The current finding also relatively granted with the reports by Molla, (2005), who detected 0.7% Salmonella isolates, 2% reported by Zubair and Ibrahim [25] and Bedaso et al. (2015) who reported 0.54% from the slaughtered goats. However, it was lower than the previous findings by Teklu and Negussie [26], who reported 11.7% in Modjo, Ethiopia and Woldemariam, et al. [27], who reported 9.8% in apparently healthy slaughtered sheep and goats in Bishoftu, Ethiopia and other reported by Wassie [22] in Addis Ababa abattoir, which was 3%. The difference in these results might be attributed to the difference in the stress condition and/or exposure of animal to the infected animal due to uncontrolled animal movement while held in the market and lairage which could lead to increased infection rate among the animal. In Zambia Sharma et al. (2001) was reported prevalence

of 2.3% from carcass swab samples of goats. In Ethiopia, Ferede et al. [23] which was 17.7% was reported from apparently healthy goats at Dire Dawa municipal abattoir, and 16.7% prevalence reported from goats slaughtered at Elfora abattoir in Ethiopia (Woldemariam [27]). The current study revealed that the carcass contaminations of the study area were low and this indicates that they were found at good hygienic condition. The result report variation might be due to the variation in the ecology of study population, sampling technique and procedure sample type and bacteriological techniques based on isolate of Salmonella or distribution of Salmonella in the study population regardless of test samples and methods of detection.

The prevalence of Salmonella on the 40 samples of knife swab was 1(0.5%) at Abyssinia export abattoirs. This result was lower than the results of Teklu and Negussie [25], who had reported 7.4% Salmonella prevalence from eviscerating knife swabs and other was reported 26.7 and 10% prevalence in two Botswana abattoirs (Motsoela et al. [28]). The variation might be associated with stress during transportation to the slaughterhouse, hygienic conditions of holding pens, processing practices, abattoir facilities and employee's hygiene and practices (Muluneh, 2019). Usually, healthy carriers' animals intermittently excrete only a few Salmonellae, unless they undergo some kind of stress (example during transport or holding in the lairages prior to slaughter). This result was in consistency with Molla et al. [29]. It is generally accepted that the carcasses of healthy slaughtered animals are free of bacteria at the time of slaughter, assuming that the animals are not in a state of exhaustion (Jay [30]). These differences in prevalence of abdominal muscle may be due to hygienic condition of abattoirs and their environment [31-42].

## Conclusion and Recommendations

In the present study, an overall of 2.5% Salmonella was isolated from 200 different types of samples examined from slaughtered goats at Abyssinia export abattoirs. The isolation of this number showed that slaughtering goats are important sources of meat contamination with the organism, and consumption of raw meat can lead to infection with zoonotic salmonellosis.

Therefore, based on the present study and findings, the following points are forwarded:

1. Application of good hygiene practice is mandatory during slaughtering processes.
2. Rational use of veterinary antimicrobials for animals is necessary to prevent subclinical diseases.
3. It should be necessary to provide public education to aware people not to consume raw meat.
4. Further investigations and studies of Salmonella up to molecular characterization should be carried out.

## References

1. (2017) Central Statistics Agency. Agricultural sample survey. Livestock and livestock characteristic report (Private Peasant Holdings). April, Addis Ababa, Ethiopia 2: 1-83.
2. Behnke RH (2010) The contribution of livestock to the economies of IGAD member states: study findings, application of the methodology in Ethiopia and recommendations for further work. IGAD 10: 1-45.
3. Endalew B, Ayalew Z (2016) Assessment of the role of livestock in Ethiopia: A Review. Am Eur J Sci Res 11: 405-410.
4. Ganteby RM (1991) Sheep: The Tropical Agriculturalist. Basingstoke and Macmillan education Limited, London, p. 1-2.
5. D'Aoust JY (1994) Salmonella and the international food trade. Int J Food Microbiol 24: 11-31.
6. (2000) Office International Des Epizootics (OIE). Manual of Standards for Diagnostic Tests and Vaccines, p. 37.
7. Tabaraie B, Sharma BK, Sharma PR, Sehgal NR, Ganguly NK (1994) Evaluation of Salmonella porins as a broad-spectrum vaccine candidate. Microbio. Immuno 38: 553-559.
8. Freeman BA (1985) Burrows Textbook of Microbiology. 22nd edn. W.B. Saunders Company, Philadelphia, London, Toronto, Mexico City, Rio de Janeiro, Sydney, Tokyo, pp. 464-472.
9. Gene O (2002) The isolation, identification and serotyping of Salmonella isolated from domestic poultry in Kars district. Kafka's Univarsikesi Veteriner Fakultesi Dergisi 8: 23-30.
10. Dworkin M, Falkow S, Rosenberg E, Schleifer K, Stackebrandt E (2006) The Prokaryotes: A Handbook on the Biology of Bacteria (3<sup>rd</sup> Edn.), p. 1.
11. Blood DC, Henderson AJ, Radosontits AJH, Gay CC (2003) A Text Book of the Diseases of Animals (9<sup>th</sup> Edn.), pp. 809-829.
12. Kadaka J, Itokazy K, Nakamura M, Taira K, Asato R (2000) An outbreak of Salmonella Weltevreden food poisoning after eating goat meat. Infect Agents Sur Rep 21: 164.
13. Carlton LD (1998) Salmonellosis: the merk veterinary manual. (8<sup>th</sup> Edn.), NJ, pp. 120-123.
14. House JK, Smith BP (1998) Current strategies for managing salmonellae infection in cattle. Veterinary medicine, pp. 756-764.
15. Pieskus J, Milius J, Michalskiene I, Zagrebneviene G (2006) He distribution of Salmonella serovars in chicken and humans in Lithuania. J Vet Med A Physiol Pathol Clin Med 53: 12-16.
16. Pepin M, Russo P, Pardon P (1997) Public health hazards from small ruminant meat products. Europe Rev Sci Technol 16(2): 415-425.
17. Sierra M, Gonzales-Fandos E, Garcia-Lopez M, Fernandez M, Prieto M (1995) Prevalence of Salmonella, Yersinia, Aeromonas, Campylobacter and cold-growing Escherichia coli on freshly dressed lamb carcasses. J Food Prot 58: 1183-1185.
18. Hansson I, Hamilton C, Ekman T, Forslund K (2000) Carcass quality in certified organic production, compared with conventional livestock production. J Vet Med B Infect Dis Vet Public Health 47: 111-120.
19. Reid C, Small A, Avery S, Buncic S (2002) Presence of foodborne pathogens on cattle hides. Food Control 13: 411-415.
20. Vatta AF (2005) Goat and Sheep keepers' veterinary manual. Onderstepoort veterinary institute, private bag underreports, South Africa 98: 26-30.

21. Quinn PJ, Carter ME, Markey BK, Carter CR (1994) *Veterinary Clinical Microbiology*, (Wolfe Publishing, London).
22. Wassie M (2004) A cross-sectional study on Salmonella in apparently healthy slaughtered sheep and goats at Addis Ababa and Modjo abattoirs, Ethiopia, MSc thesis, Addis Ababa University, Faculty of Veterinary Medicine, Bishoftu, Ethiopia.
23. D'Aoust JY (1989) Salmonellae In: Doyle M (9<sup>th</sup> Edn.), *Food borne bacterial pathogens*. Marcel Dekker Inc New York, pp. 328-413.
24. Ferede B, Desissa F, Feleke A, Tadesse G, Moje N (2015) Prevalence and antimicrobial susceptibility of Salmonella isolates from apparently healthy slaughtered goats at Dire Dawa municipality abattoir, Ethiopia. *J Microbial Antimicrob* 7(1): 1-5.
25. Zubair AI, Ibrahim KS (2012) Isolation of Salmonella from slaughtered animals and sewage at Zakho abattoir, Kurdistan Region, Iraq. *Res Opin Anim Vet Sci* 3(1): 20-24.
26. Teklu A, Negussie H (2011) Assessment of risk factors and prevalence of Salmonella in slaughtered small ruminants and environment in an export abattoir, Modjo, Ethiopia. *American-Eurasian J Agric Environ Sci* 10(6): 992-999.
27. Woldemariam E, Molla D, Alemayehu, Muckle A (2005) Prevalence and distribution of salmonella in apparently healthy slaughtered sheep and goats in Debrezeit, Ethiopia. *Small Rumin Res* 58: 19-24.
28. Woldemariam E (2003) Prevalence and distribution of Salmonella in apparently healthy slaughtered sheep and goats in Debre Zeit, Ethiopia. AAU, FVM, Debre Zeit, Ethiopia, DVM. Thesis.
29. Motsoela C, Collison EK, Gashe BA (2002) Prevalence of salmonella in two Botswana abattoir environments. *J Food Prot* 65: 1869-1872.
30. Molla W, Molla B, Alemayehu D, Muckle A, Cole L, et al. (2006) Occurrence and antimicrobial resistance of Salmonella serovars in apparently healthy slaughtered sheep and goats of central Ethiopia. *Trop Anim Health Prod* 38: 455-462.
31. Jay JM (2000) *Modern Food Microbiology*. 6th ed. Maryland: Aspen publication, pp. 511-525.
32. (2007) Domestic animal genetic resources of Ethiopia. IBC (Institute of Biodiversity Conservation), Addis Ababa, Ethiopia.
33. Arruda SGB, Biscontini TMB, Stamford TLM (2004) Microbiological characterization of goat meat subjected to different forms of management. *Hygiene Alimentary* 18: 58-62.
34. Alemayehu D, Molla B, Muckle A (2003) Prevalence and antimicrobial resistance pattern of Salmonella isolates from apparently healthy slaughtered cattle in Ethiopia. *Tropical Animal Health and Production* 35: 309-319.
35. Behnke RH (2010) The contribution of livestock to the economies of IGAD member states: study findings, application of the methodology in Ethiopia and recommendations for further work. *IGAD* 10: 1-45.
36. Cooke EM, Todd E (1990) Epidemiology of foodborne illness: UK. *Lancet* 336: 790-793.
37. Gould LH, Mungai E, Behravesh CB (2014) Outbreaks attributed to cheese: differences between outbreaks caused by unpasteurized and pasteurized dairy products, United States, 1998-2011. *Foodborne Pathog Dis* 11: 545-551.
38. (1998) International Organization for Standardization (ISO). ISO 6579: Microbiology of Food and Animal Feeding Stuff— Horizontal Method for the Detection of Salmonella, (ISO, Geneva).
39. Molla B, Mohammed A, Salah W (2004) Salmonella prevalence and distribution of serotypes in apparently healthy slaughtered camels (*Camelus dromedarius*) in Eastern Ethiopia. *Tropical Animal Health and Production* 36(5): 451-458.
40. Quinn PJ, Markey BK (2003) Concise review of veterinary microbiology. Back well publishing, p. 6.
41. Rabsch W, Altier C, Tschape H, Baumler AJ (2003) Food borne Salmonella infection. In: Torrence ME, Isaacson RE (Eds.), *Microbial food safety in animal agriculture*. Blackwell Publishing. USA, pp. 97-108.
42. W Molla, B Molla, D Alemayehu, A Muckle, L Cole, et al. (2005) Prevalence and distribution of Salmonella in apparently healthy slaughtered Occurrence and antimicrobial resistance of Salmonella serovars in apparently healthy slaughtered sheep and goats of central Ethiopia.

ISSN: 2574-1241

DOI: 10.26717/BJSTR.2023.51.008053

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